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Asymmetric Bridging Annulation Reaction Involving the Intramolecular Conjugate Addition of Chiral Imines to Enoates: Access to a Polycyclic System Related to the Taxane Core

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Abstract: (R)-1-phenylethylamine-induced cyclization of ketoenoate 24 led to a 2:1 mixture of "all-cis" polycyclic adducts 25 and 26, structurally related to the taxane series.

Ten years ago, we reported that chiral imines 3, derived from *racemic* 2-alkylcyclanones 1 and optically active 1-phenylethylamine 2, add, *under neutral conditions*, to electrophilic alkenes 4 leading, after hydrolytic work-up, to 2,2-dialkylcyclanones 5 with a good yield and excellent regio- and stereoselectivity.²

Ph. Me. H. NH₂ Ph. Me. H. NH₂
$$i: EWG$$

$$\frac{2}{i: H_3O^+}$$

$$\frac{4}{i: H_3O^+}$$

$$5$$

(typical ee: 90-98 %)

This Michael addition, which tolerates a great variation in the nature of the starting cyclanone and the electrophilic alkene, constitutes indisputably one of the most potent methodology for the enantioselective elaboration of quaternary carbon centers. In this respect, it has been successfully applied by ourselves and others to the asymmetric synthesis of various compounds of natural origin, including terpenes, steroids and alkaloids.² The mechanistic aspects of this reaction have also been extensively investigated, from both experimental and theoretical viewpoints.² Several intranolecular variants of this Michael addition have been developed; thus, for example, chiral imines 6^3 and 8^4 underwent a facile carbocyclization under thermal conditions, furnishing, after hydrolytic work-up, derivatives 7 and 9, respectively, with an excellent enantioselectivity (α 90%).

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On the other hand, (R)-1-phenylethylamine-induced *spiroannulation* of ketoenoate 10 led to ketoester (15, 2R)-11 with a high degree of control ($\geq 90\%$) of the two newly created stereogenic centers.⁵ In a similar fashion, *bridging annulation* of ketoenoate 12, induced by (ent)-2, gave the bicyclo[3.3.1]nonane derivative 13 with a high level of stereoselectivity (100% de, 90% ee).⁶

However, this bridging annulation proved to be very sensitive to the size of the newly created ring; thus, the [3.2.1] and [4.3.1] bicyclic compounds 15 and 17, elaborated from ketoenoates 14 and 16, respectively, were both obtained with a low degree of stereoselectivity (60% de, 30% ee and 20% de, respectively).⁶

In connection with our synthetic efforts in the taxane series (exemplified by taxol 18)⁷, we recently planed the construction of the [AB] ring framework of these systems, by extending the present bridging annulation. However, all attempts at elaborating the bicyclo[5.3.1]undecane derivative 20 by 1-phenylethylamine-induced cyclization of ketoenoate 19 were invariably unsuccessful, a reflect of the difficulties usually encountered in establishing eight-membered carbocycles from open-chain precursors.⁶

In light of the preceding observation, we reasoned that the intercalation of an aromatic moiety, precursor of the future C ring of taxanes, in the side-chain of 19, by reducing significantly the degrees of freedom of the system, as well as the transannular strain in the eight-membered adduct, would favor the annulation reaction.⁸ This was tested on model ketoenoate 24, efficiently elaborated by condensing first the lithio derivative of ester 21⁹ with bromomethylsafrole 22¹⁰ (i: 21, 1.5 eq of LDA, THF, 30 min at -78 °C; ii: 22, -78 °C \rightarrow 20 °C, then 12 h at this temperature; iii: 1N HCl, 1 h at 20 °C). Compound 23, thus obtained with a 58% overall yield, was

then converted into ketoenoate 24^{11} through a two-step sequence ($i : cat \, OsO_4$, NaIO₄, Et₂O, H₂O, 1 h at 20 °C; $ii : Ph_3P=CH-COOMe$, CH₂Cl₂, 24 h at 20 °C, 70 % overall yield).

Ketoenoate 24 was then cyclized [2 eq of (R)-1-phenylethylamine, 72 h in refluxing toluene, with removal of water], leading with a 48 % yield to a 2:1 mixture of isomeric adducts 25¹² and 26¹³, respectively. These were separated by preparative HPLC¹⁴ and fully characterized. The same "all-cis" relative stereochemistry in the tricyclic systems of 25 and 26 was established through the complete assignments of the ¹H and ¹³C NMR resonances, including 1D and 2D experiments, PFG-phase sensitive DQF COSY, and ¹H-¹³C direct and long range correlations (PFG-HMQC and PFG-HMBC). The proposed absolute configurations of the three newly created stereogenic centers in 25 and 26, although not definitively established, rests on the mechanism of the present Michael addition, assuming that the addition took place predominantly on the less hindered side of the intermediary chiral imine, anti to the phenyl group of the inducer.^{2,6}

It is clear that the formation of adducts 25 and 26 from 24 involves the intermediary Michael adducts $(e.g.\ 27)$ which underwent a lactamization side-reaction. In order to minimize such a lactamization process, 1-phenylethylamine-induced cyclization of ketoenoate 28, the *tert*-butyl ester analog of 24, was next investigated. However, when 28 was submitted to the aforementioned cyclization conditions, only the β , γ -ethylenic ester 29 was isolated.

An efficient access of a polycyclic system related to the taxane core has thus been achieved. New synthetic developments of this methodology are currently under investigation in our laboratory.

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- 24: oil; ¹H NMR (200 MHz, CDCl₃) & 6.93 (dt, J = 15.2 Hz, J = 6.1 Hz, 1H) 6.50 (s, 1H) 6.45 (s, 1H) 5.86(s, 2H) 5.58(dt, J = 15.2 Hz, J = 3.0 Hz, 1H) 3.67 (s, 3H) 3.64 (s, 3H) 3.35 (dd, J = 6.1 Hz, J = 3.0 Hz, 1H) 2.75 (s, 2H) 2.5-2.2 (m, 6H) 1.7-1.6 (m, 2H); ¹³C NMR (50 MHz, CDCl₃) & 210.4 (C), 177.5 (2 C), 147.2 (C), 146.9 (CH), 129.6 (C), 127.6 (2 C), 122.0 (CH), 110.7 (CH), 110.0 (CH), 101.1 (CH₂), 56.2 (CH₃), 52.1(CH₃), 48.4 (C), 41.7 (CH₂), 38.5 (2 CH₂), 35.4 (CH₂), 33.7 (2 11
- **25**: solid, mp 210-212 °C (MeOH); $[\alpha]D^{20}$ + 49.1 (c 1.7, CCl4); MS (70 eV): 459 (M⁺), 355, 296, 206, 175, 149, 105; IR (CHCl₃): 1729, 1660, 1633 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.20 (m, 5H), 6.65 (s, 1H), 6.43 (q, 1H, J = 7.1 Hz), 6.40 (s, 1H,), 5.90 (s, 2H), 4.99 (dt, 1H, 1H, 1Hz, 1Hz), 3.69 (s, 3H), 3.04 (dd, 1H, J = 13.5 Hz, J = 13.2 Hz), 2.95 (d, 1H, J = 14.5 Hz), 2.92 (dd, 1H, J = 14.5 Hz), 2. = 17.9 Hz, J = 7.0 Hz), 2.83 (d, 1H, J = 14.6 Hz), 2.75 (ddd, 1H, J = 10.0 Hz, J = 2.0 Hz, J = 2.0 Hz), 2.66 (d, 1H, J = 17.9 Hz), 2.44 (d, 1H, J = 13.5 Hz), 2.30 (ddd, 1H, J = 16.5 Hz, J = 2.0 Hz J = 2.0 HzHz), 2.15 (ddd, 1H, J = 16.5 Hz, J = 7.0 Hz, J = 3.0 Hz), 2.04 (dd, 1H, J = 14.1 Hz, J = 10.0 Hz), 2,00 (dd, 1H, J = 13.2 Hz, J = 7.0 Hz), 1.71 (d, 3H, J = 7.1 Hz), 1.50 (dd, 1H, J = 14.9 Hz, J = 3.0 Hz); 13 C NMR (50 MHz, CDCl₃) δ 176.0 (C), 168.5 (C), 146.7 (2 C), 141.8 (C), 134.2 (C), 133.4 (C), 129.9 (C), 128.7 (2 CH), 126.5 (CH), 125.4 (2 CH), 110.0 (CH), 109.7 (CH), 107.7 (CH), 100.9 (CH₂), 52.1 (CH₃), 49.9 (CH), 45.7 (C), 42.8 (CH₂), 39.5 (CH), 37.8 (CH₂), 37.5 (CH₂), 36.1 (CH),
- 33.8 (CH2), 30.0 (CH2), 15.5 (CH3).
 26: amorphous solid; $[\alpha]D^{20} + 54.0$ (c 3.6, CCl4); MS (70 eV): 459 (M⁺), 355, 296, 206, 175, 149, 105; IR (CHCl3): 1729, 1660, 1633 cm⁻¹; ¹H NMR (400 MHz, CDCl3) δ 7.35-7.20 (m, 5H), 6.65 (s, 1H), 6.42 (s, 1H), 6.10 (q, 1H, J = 7.0 Hz), 5.90 (s, 2H), 5.04 (ddd, 1H, J = 5.0 Hz, J = 2.5 Hz, J = 2.0 Hz), 3.68 (s, 3H), 3.16 (d, 1H, J = 14.0 Hz), 3.00 (dd, 1H, J = 17.0 Hz, J = 7.0 Hz), 2.97 (d, 1H, J = 17.0 Hz), 2.97 (d, 1H, J = 17.0 Hz), 2.97 (d, 1H, J = 17.0 Hz), 2.98 (d, 1H, J = 17. = 14.0 Hz, 2.94 (dd, 1H, J = <math>14.0 Hz, J = 6.0 Hz), 2.74 (dd, 1H, J = <math>2.5 Hz, J = 2.0 Hz), 2.61 (d, 1H, J = <math>2.5 Hz), 2.61 (d, 2L)1H, J = 17.0 Hz, 2.38 (d, 1H, J = 14.0 Hz), 2.24 (ddd, 1H, J = 17.0 Hz, J = 5.0 Hz, J = 3.0 Hz), 2.12 (ddd, 1H, J = 17.0 Hz, J = 2.5 Hz, J = 2.5 Hz), 2.02-1.95 (m, 2H), 1.82 (d, 3H, J = 7.0 Hz), 1.50 (dd, 1H, J = 15.0 Hz, J = 3.0 Hz); 13 C NMR (50 MHz, CDCl₃) δ 176.5 (C), 167.6 (C), 146.0 (2 C), 141.0 (C), 134.2 (C), 133.8 (C), 128.8 (C), 128.2 (2 CH), 126.2 (CH), 125.7 (2 CH), 109.8 (CH), 109.3 (CH), 106.7 (CH), 100.0 (CH₂), 51.8 (CH₃), 51.7 (CH), 45.0 (C), 42.7 (CH₂), 39.1 (CH), 37.4 (CH₂), 36.9 (CH₂), 36.0 (CH), 33.6 (CH₂), 29.7 (CH₂), 17.2 (CH₃)
- Analytical HPLC: Hichrom column (Spherisorb S 5 W), lenght: 25 cm, internal diameter: 4.9 mm; detection: UV at 254 nm; eluent: cyclohexane/AcOEt 90:10; flow rate: 2 ml min⁻¹; retention times: 25: 7.79 min, **26**: 6.83 min.
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